

OECD QSAR Toolbox v.4.1

Example illustrating endpoint vs. endpoint correlation for apical endpoints

Outlook

- **Background**
- Objectives
- The exercise
- Workflow

Background

This presentation is designed to introduce the user with:

- Illustration of different types endpoint vs. endpoint correlations using:
 - LLNA and GPMT skin sensitization data
 - DPRA and LLNA skin sensitization data
 - Skin sensitization and Ames mutagenicity data

Outlook

- Background
- **Objectives**
- The exercise
- Workflow

Objectives

This presentation demonstrates a number of functionalities of the Toolbox:

- Illustration of endpoint vs. endpoint correlations using different type endpoint data

Outlook

- Background
- Objectives
- **The exercise**
- Workflow

The exercise

- Illustration of different endpoint data correlations:
 - LLNA vs. GPMT skin sensitization data
 - DPRA (reactivity) vs. LLNA (skin sensitization) data
 - GPMT (skin sensitization) vs. Ames mutagenicity data

Outlook

- Background
- Objectives
- The exercise
- **Workflow**

Workflow

- **The Toolbox has six modules which are typically used in a workflow:**
 - Chemical Input
 - Profiling
 - Endpoints
 - Category Definition
 - Filling Data Gaps
 - Report
- **In this example we will use the modules in a different order, tailored to the aims of the example.**

Outlook

- Background
- Objectives
- The exercise
- **Workflow**
 - **Correlation of data - background**

Correlation of endpoint data

Background

- This functionality introduce the user with opportunity to analyze correlations between selected gap filling endpoint (endpoint used for prediction) and other endpoint data.
- It is applicable for correlation analysis of data presented in ordinary, interval or ratio scale.
- If correlated data are measured in interval or ratio scale they are transformed in ordinary scale and the strength of the correlation is estimated by Spearman correlation coefficient.
- Basically, this functionality provides a correlation between target endpoint (this is the initial endpoint selected by the user) displayed on ordinate axis (Y-axis) and other endpoint data displayed on abscissa (X-axis).

Correlation of endpoint data

Spearman coefficient factor

- Spearman's rank correlation coefficient is a nonparametric rank statistic proposed by Charles Spearman as a measure of the strength of an association between two variables. It assesses how well the relationship between two variables can be described using a monotonic function.
- Spearman correlation coefficient could be used for exploring the covary between:
 - two ranked variables
 - one measurement variable and one ranked variable (in this case, the measurement variable need to be to converted to ranks)
- Spearman correlation varies from -1 to +1 and the interpretation of the coefficient factor is provided below:
 - 0.00 – 0.19 – very weak correlation
 - 0.20 – 0.39 – weak correlation
 - 0.40 – 0.59 – moderate correlation
 - 0.60 – 0.79 – strong correlation
 - 0.80 – 1.0 – very strong

Outlook

- Background
- Objectives
- The exercise
- **Workflow**
 - Correlation of data – background
 - **Types endpoint correlations**

Types endpoint correlations

Types endpoint correlations are as follows:

- Continuous vs. continuous*
- Categorical vs. categorical:
 - ✓ Categorical vs. categorical
 - ✓ Categorized continuous vs. categorical
 - ✓ Categorized continuous vs. categorized continuous*

*Both type correlation is not illustrated in this presentations. They are presented in "Tutorial_4_TB 4.1_Illustrating endpoint vs. endpoint correlation using ToxCast data"

Outlook

- Background
- Objectives
- The exercise
- **Workflow**
 - Correlation of data – background
 - **Types endpoint correlations**
 - Categorical vs. categorical

Types endpoint correlations

Categorical vs. categorical

- The aim of this type correlation is to illustrate how categorical type data correlates each other.
- Categorical type data is the statistical data type consisting of categorical variables or of data that has been converted into that form. Such data is binary Ames data (dichotomic type): positive, negative or polytomic type data such as GPMT data: strong, weak and negative.
- Two examples illustrating this type correlation will be demonstrated:
 - Example 1: Correlation of two types skin sensitization data
 - LLNA (Positive, Negative) vs. GPMT (Weakly positive, Strongly positive, Negative)
 - Example 2: Correlation of skin sensitization and Ames mutagenicity data
 - LLNA (Negative, Weakly positive, Strongly positive) vs. AMES (Positive, Equivocal, Negative)
- Step by step workflow is presented on next few slides. Summary of the workflow steps are provided below:
 - *Load Skin sensitization database (step 1)*
 - *Gather experimental data (step 2)*
 - *Define target endpoint (step 3)*
 - *Enter Gap filling (step 4)*
 - *Perform correlation between endpoints (step 5).*

Types endpoint correlations

Categorical vs. categorical

Load Skin sensitization database – step 1

Example 1: Correlation of LLNA and GPMT data

The screenshot shows the QSAR Toolbox software interface. The top menu bar includes 'Input', 'Profiling', 'Data', 'Category definition', 'Data Gap Filling', and 'Report'. The toolbar below contains icons for 'New', 'Open', 'Close', 'Name', 'Structure', 'Composition', 'Select', 'Delete', 'Chemical', 'Database', 'Inventory', 'List', 'Substructure (SMARTS)', 'Query', and 'Define'. The left sidebar shows a document list with 'ToxCastDB' and 'Skin Sensitization'. The main workspace displays a 'Chemical List' table with columns for 'Filter endpoint', 'Structure', and 'Inventory'. A 'Select database' dialog box is open, listing various databases, with 'Skin Sensitization' highlighted. Numbered callouts (1-5) indicate the steps: 1. Click 'Input' in the menu; 2. Click 'Database' in the toolbar; 3. Select 'Skin Sensitization' in the dialog; 4. Click 'OK' in the dialog; 5. Chemical structures are loaded into the table.

1. **Go** to "Input";
2. **Click** "Database" button;
3. **Select** "Skin sensitization" database;
4. **Click** OK;
5. The chemicals from database have been loaded on datamatrix;

Types endpoint correlations

Categorical vs. categorical

Gather experimental data – step 2

Example 1: Correlation of LLNA and GPMT data

The screenshot displays the QSAR Toolbox interface. The top menu bar includes 'Data', 'Profiling', 'Data', 'Category definition', 'Data Gap Filling', and 'Report'. The 'Data' menu is open, showing 'Gather' (callout 3) and 'IUCLID6'. The 'Documents' pane shows 'Document 1' with 'ToxCastDB' and 'Skin Sensitization' selected. The 'Options' pane shows 'Skin Sensitization' checked under 'Rodent Inhalation Toxicity' (callout 2). The 'Filter endpoint tree...' pane lists various endpoints, with 'Skin Sensitization' highlighted. The 'Read data?' dialog box is open, showing 'All endpoints' selected and 'OK' button (callout 4). A message box at the bottom right states '2019 points added across 1201 chemicals.' with an 'OK' button (callout 5).

1. Go to "Data";
2. Select "Skin sensitization";
3. Click "Gather"
4. Click "OK"
5. Click "OK";

What is “scale” and “scale conversion” ?

Reminder slide

- Skin sensitisation as an example is a “qualitative” endpoint for which the results are presented with categorical type of data (for example: positive; negative; weak sensitizer; strong sensitizer, etc).
- Skin sensitisation potential of the chemicals came from different authors coded with different names (for example: data from John Moores University of Liverpool are: *Strongly sensitizing, Moderately sensitizing etc.*; data from European centre for Ecotoxicology and Toxicology of chemicals are: *Positive, Negative, and Equivocal*).
- The main purpose of the scales is to unify all data available in the Toolbox databases for a certain endpoint.
- “Scale conversion” is the TB instrument to create conversions between scales. More reasonable is to convert more informative to less informative scale.
- The default scale for Skin Sensitisation data is “Skin Sensitisation ECETOC”. It converts all skin sensitization data into: Positive and Negative. This allows skin sensitization data to be used as much as possible for gap filling purposes.

Types endpoint correlations

Categorical vs. categorical

Define target endpoint – step 3

Example 1: Correlation of LLNA and GPMT data

The screenshot shows the QSAR Toolbox interface. On the left, the 'Filter endpoint tree...' is expanded to show 'Sensitisation' > 'In Vivo' > 'LLNA' selected. On the right, a table displays data for various endpoints across six columns. A red box highlights the cell for 'LLNA' in column 1, and a blue callout bubble with the number '1' points to it.

Endpoint	1	2	3	4	5	6
Structure	<chem>Oc1ccc(O)cc1</chem>	<chem>CCCCCCCC</chem>	<chem>NC(=O)CC(=O)N</chem>	<chem>Nc1ccccc1</chem>	<chem>CCCCCCCC</chem>	<chem>CCCCCCCC</chem>
Structure info						
Parameters						
Physical Chemical Properties						
Environmental Fate and Transport						
Ecotoxicological Information						
Human Health Hazards						
Acute Toxicity						
Bioaccumulation						
Carcinogenicity						
Developmental Toxicity / Teratogenicity						
Genetic Toxicity						
Immunotoxicity						
Irritation / Corrosion						
Neurotoxicity						
Photoinduced toxicity						
Repeated Dose Toxicity						
Sensitisation						
Skin						
In Vivo						
GPMT	(332/333)					
LLNA	(614/727)					
EC3		M: Negative	M: Positive	M: Negative		M: Positive
Miscellaneous	(419/607)	M: Negative				
Undefined Assay	(1/1)					
ToxCast						
Toxicity to Reproduction						
Toxicokinetics, Metabolism and Distribution						

The target endpoint is EC3 data associated with LLNA assay.

1. **Click** on the cell associated with target endpoint;

Types endpoint correlations

Categorical vs. categorical
Define target endpoint – step 3

Example 1: Correlation of LLNA and GPMT data

The screenshot shows the QSAR Toolbox interface with the following components:

- Top Bar:** QSAR TOOLBOX logo and navigation icons for Input, Profiling, Data, Category definition, Data Gap Filling, and Report.
- Workflow Bar:** Gap Filling and Workflow tabs.
- Documents Panel:** A tree view showing 'Document 1' > 'ToxCastDB' > 'Skin Sensitization'. The 'EC3' filter box is highlighted with a red box and a callout '1'.
- Human Health Hazards Table:** A table with columns 1-8 and rows for 'Structure', 'Sensitisation', 'In Vivo', and 'LLNA'. The 'LLNA' row is expanded to show 'EC3' with a value of '(614/727)'. The cell for 'M: Negative' is highlighted with a blue box and a callout '2'.

	1	2	3	4	5	6	7	8
Structure	<chem>Oc1ccc(O)cc1</chem>	<chem>CCCCCCCC</chem>	<chem>CC(O)CC(=O)O</chem>	<chem>Nc1ccccc1</chem>	<chem>CC1(C)CC2(C)CC3(C)CC4(C)CC5(C)CC6(C)CC7(C)CC8(C)CC9(C)CC10(C)CC23456789</chem>			
Human Health Hazards								
Sensitisation								
In Vivo								
LLNA								
EC3	(614/727)	M: Negative	M: Positive			M: Positive	M: Positive	M: Positive

- 1. Insert** type "EC3" data associated with LLNA assay in the filter box, then **press** "Enter" and automatically opening the tree to the target endpoint;
- 2. Click** on the cell associated with target endpoint;

Types endpoint correlations

Categorical vs. categorical

Enter Gap filling – step 4

Example 1: Correlation of LLNA and GPMT data

The screenshot shows the QSAR Toolbox software interface. The top menu bar has 'Data Gap Filling' selected. The left sidebar shows 'Documents' with 'Skin Sensitization' selected. The main workspace displays a chemical structure and a data table with columns 1-5. A dialog box titled 'Possible data inconsistency' is open, showing 'Native scale/unit' with 'Skin sensitization EC3(ratio)' checked. The 'Gap filling scale/unit' section has 'Skin sensitization II (ECETOC)' selected. The dialog also shows 'converted data' and 'Data 727/727; Chemicals 614/614'. Red callout boxes with numbers 1, 2, 3, and 4 highlight specific steps in the workflow.

Note: By default EC3 data has been converted into binary categories: positive/negative based on scale "Skin sensitization II (ECETOC)". For the purpose of this exercise, Skin sensitization I (OASIS) will be used. This scale converts EC3 data into three categories: Strongly positive (EC3 0-10%), Weakly positive (EC3 10-50%) and Negative (EC3>50%).

Enter Gap filling and apply read across. Read across is applied because a categorical type data is analyzed. Follow the steps:

1. **Go** to "Data Gap filling";
2. **Select** "Read-across";
3. **Select** "Skin sensitization II (ECETOC)" scale (see Note);
4. **Click** "OK";

Types endpoint correlations

Categorical vs. categorical

Enter Gap filling – step 4

Example 1: Correlation of LLNA and GPMT data

The screenshot shows the QSAR Toolbox interface during the 'Data Gap Filling' step. The main window displays a 'Filter endpoint tree' on the left and a data table on the right. The table has columns for target endpoints (1-15) and rows for chemical structures. The bottom row shows correlation results: 'M: Negative', 'M: Positive', 'M: Positive', 'M: Positive', 'M: Positive', 'M: Positive', 'M: Negative', 'M: Positive', 'M: Positive', 'M: Positive'. An 'Information' dialog box is open in the foreground, displaying the message: '13 observed values for 12 chemicals were excluded due to missing X descriptor value(s)'. A red number '1' is placed next to the 'OK' button in the dialog box.

The message informing the user for how many chemicals with experimental data are excluded from gap filling due to missing X-descriptor value appeared 1. **Click “OK”;**

Types endpoint correlations

Categorical vs. categorical

Perform correlation between LLNA and GPMT data– step 5

Example 1: Correlation of LLNA and GPMT data

The screenshot displays the QSAR Toolbox interface. On the left, the 'Documents' panel shows 'Document 1' with a sub-entry 'Enter GFR(A) with 602 chemicals, 713 data points'. Below it, the 'Data Gap Filling Settings' panel has checkboxes for 'Only endpoint relevant' and 'Only chemical relevant', and a section 'At this position:' with a list of workflow types. The main workspace features a 'Filter endpoint tree...' on the left, a 'Structure' panel with a chemical structure, and a data table with columns for chemical IDs and predicted values. A 'Choose one' dialog box is open, showing a list of calculation options: All, Mode, Lowest mode, Highest mode, Median, Lower median, Higher median, Minimal, and Maximal (selected). Callouts 1, 2, 3, and 4 point to the 'Calculation options' menu, the 'Data usage' section, the 'Maximal' option, and the 'OK' button, respectively. The right sidebar contains various settings like 'Select / filter data', 'Gap filling approach', and 'Prediction approach options'.

Correlation assumes a single value per chemical to be used. In this respect the default calculation settings should be changed from "All" to something different. In our case study we play a worst case scenario, thus an option "All" is changed to "Maximal" values. Follow the steps:

1. **Open** "Calculation options";
2. **Click** on "Data usage" menu item;
3. **Select** Maximal;
4. **Click** "OK";

Types endpoint correlations

Categorical vs. categorical

Perform correlation between LLNA and GPMT data– step 5

Example 1: Correlation of LLNA and GPMT data

The screenshot displays the QSAR Toolbox software interface. The main window shows a 'Filter endpoint tree...' window with a tree structure under 'Human Health Hazards' > 'Sensitisation' > 'Skin'. The 'GPMT (81/82)' node is selected and circled in red (callout 4). A 'Select endpoint descriptor' dialog is open, showing 'SMWN (81/82)' selected (callout 3). A 'Possible data inconsistency' dialog is also open, showing 'Skin sensitisation I (Oasis)' selected (callout 6) and '82 from scale Skin sensitisation IV (GPMT)' as converted data (callout 7). The 'Data Gap Filling Settings' panel is visible on the left, with 'Only endpoint relevant' and 'Only chemical relevant' checked. A scatter plot at the bottom shows the correlation between log Kow (X-axis) and ECI (Y-axis), with a single data point at approximately (14, 82) (callout 5). The 'Descriptor/data' tab is selected in the bottom right (callout 1), and the 'Select / filter data' button is highlighted (callout 2).

1. Open Descriptor/data tab;
2. Click on Select endpoint tree descriptor;
3. **Open** nodes under "Sensitization" node;
4. **Select** second endpoint, which will be placed on X-axis circled in red box: SMWN;
5. **Click** "OK" button;
6. **Select** Scale I OASIS
7. **Click** OK

Types endpoint correlations

Categorical vs. categorical

Perform correlation between LLNA and GPMT data– step 5

Example 1: Correlation of LLNA and GPMT data

The message inform the user for how many chemicals will be excluded from correlation due to missing data for SMWN endpoint appears. This will not affect the value of correlation coefficient 1. **Click “OK”**;

Types endpoint correlations

Categorical vs. categorical

Perform correlation between LLNA and GPMT data– step 5

Example 1: Correlation of LLNA and GPMT data

The screenshot displays the QSAR Toolbox interface during a correlation analysis. The 'Filter endpoint tree...' panel shows a hierarchy: Structure > Human Health Hazards > Sensitisation > In Vivo > LLNA > EC3. The 'Data Gap Filling' table shows 10 columns of chemical structures with corresponding 'M: Positive' or 'M: Negative' values. The 'Prediction' chart shows a Spearman coefficient of 0.53 (moderate) and a bar chart of EC3 values for Negative, Weakly positive SMWN, and Strongly positive categories. A dialog box 'Edit calculator options' is open, showing 'Maximal' selected for the calculation mode. Numbered callouts (1-4) highlight key steps: 1. Select 'Descriptor/ data' in the right sidebar; 2. Click 'Edit descriptor options'; 3. Select 'Maximal' in the dialog; 4. Click 'OK' in the dialog.

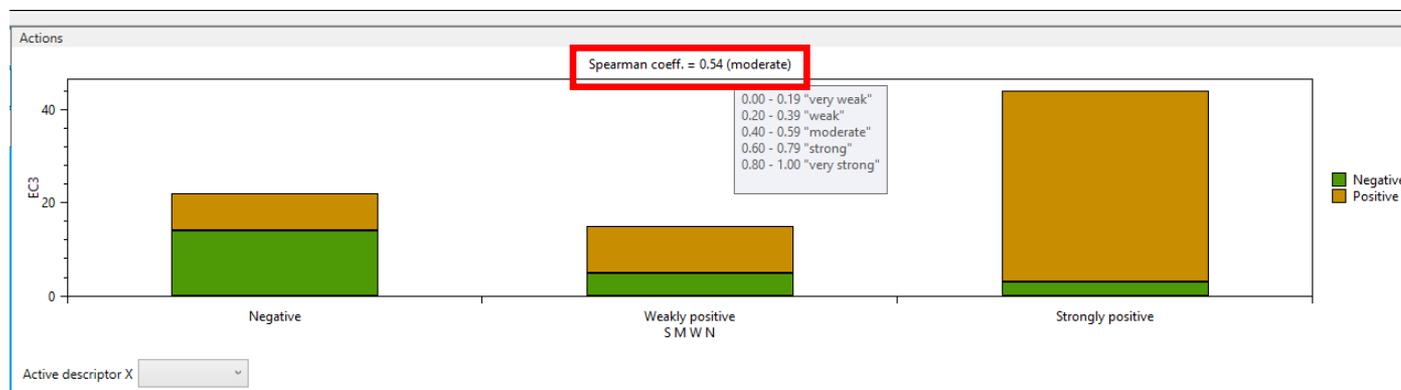
1. **Select** "Descriptor/ data"; 2. **Click** "Edit descriptor options" ; 3. **Select** "Maximal"; 4. **Click** OK

Types endpoint correlations

Categorical vs. categorical

Interpretation of correlation results (LLNA vs. GPMT)

Example 1: Correlation of LLNA and GPMT data



- Correlation analysis between two categorical type skin sensitization data (LLNA and GPMT) shows moderate endpoint correlation (Spearman coefficient is 0.54).

Types endpoint correlations

Categorical vs. categorical

- The second example illustrating categorical vs. categorical type correlation is:
 - Example 2: Correlation of Skin sensitization and Ames mutagenicity data
 - LLNA (Negative, Weakly positive, Strongly positive)
 - AMES (Positive, Equivocal, Negative)
- Step by step workflow is presented on next few slides. Summary of the workflow steps are provided below:
 - *Load Skin sensitization database (step 1) – skipped, because this database is already loaded on data matrix*
 - *Gather experimental data (step 2)*
 - *Define target endpoint (step 3)*
 - *Enter Gap filling (step 4)*
 - *Perform correlation between endpoints (step 5)*

Types endpoint correlations

Categorical vs. categorical
Gather experimental data – step 2
 Sidebar of database relevancy

Once the endpoint is selected, the relevant databases are highlighted.

The screenshot displays the QSAR Toolbox interface. At the top, there is a navigation bar with icons for Input, Profiling, Data, Category definition, Data Gap Filling, and Report. Below this is a menu bar with Data, Import, and Export options. The main workspace is divided into several panels:

- Documents:** Shows a tree view for 'Skin Sensitization' with 602 chemicals.
- Filter endpoint tree...:** A hierarchical tree of endpoints. A 'Legend' dialog box is open over it, showing 'Target endpoint' with a green square for 'Have data for target endpoint' and a white square for 'Have no data for target endpoint'.
- Options:** A panel on the left with 'Group by: Category', 'Sort by: Name', and 'Color by: Endpoint sele'.
- Inventories:** A list of databases on the left, with 'Genotoxicity OASIS' and 'ECHA CHEM' highlighted in green.
- Data Table:** A table with 6 columns. The first column contains chemical structures. The second column contains assay names and counts. The third column contains assay results (e.g., 'M: Negative').

Assay	Count	Result	Result	Result	Result
Bacterial Reverse Mutation Assay (e.g. Ames ...)					
Gene mutation					
Salmonella typhimurium					
No S9 Info	(363/431)				
With S9	(434/3174)	M: Negative			
Without S9	(446/3278)	M: Negative			
In Vitro Mammalian Cell Micronucleus Test (18/19)					
In Vitro Mammalian Chromosome Aberra(170/305)					
Mammalian Cell Gene Mutation Assay (56/56)					
In Vivo (88/141)					
Immunotoxicity					
Irritation / Corrosion					
Neurotoxicity					
Photoinduced toxicity					
Repeated Dose Toxicity					
Sensitisation AW SW AOP (1201/2019)		M: Negative	M: Negative	M: Positive	M: Negative
ToxCast					
Toxicity to Reproduction					
Toxicokinetics, Metabolism and Distribution					

Types endpoint correlations

Categorical vs. categorical

Gather experimental data – step 2

Example 2: Correlation of LLNA and AMES data

The screenshot shows the QSAR Toolbox interface. On the left, the 'Documents' tree has 'Skin Sensitization' selected under 'Document 2', highlighted with a red box and a callout '1'. The 'Data' tab is selected in the top navigation bar, highlighted with a callout '2'. The main window displays a table of chemical structures and their correlation results for 'Sensitisation' (LLNA and EC3). The table has 9 columns for different chemical structures and rows for 'Sensitisation' and 'EC3'. The 'Sensitisation' row shows 'M: Negative' for structure 2, 'M: Positive' for structures 3, 7, 8, and 9. The 'EC3' row shows 'M: Negative' for structure 2, 'M: Positive' for structures 3, 7, 8, and 9.

EC3	1	2	3	4	5	6	7	8	9
Structure	<chem>Oc1ccc(O)cc1</chem>	<chem>CCCCCCCC</chem>	<chem>CC(C)C(=O)C</chem>	<chem>Nc1ccccc1</chem>	<chem>CC1(C)CC(C)CC1</chem>	<chem>CC1(C)CC(C)CC1</chem>	<chem>CC1(C)CC(C)CC1</chem>	<chem>CC1(C)CC(C)CC1</chem>	<chem>CC1(C)CC(C)CC1</chem>
Human Health Hazards									
Sensitisation									
Skin									
In Vivo									
LLNA									
EC3	(614/727)	M: Negative	M: Positive				M: Positive	M: Positive	M: Positive

1. In order to start with next example please **click** on the level Skin sensitization from document tree 2. **Click** on Data tab

Types endpoint correlations

Categorical vs. categorical

Gather experimental data – step 2

Example 2: Correlation of LLNA and AMES data

The screenshot shows the QSAR Toolbox interface. The top menu bar includes 'Data', 'Import', and 'Export'. The 'Filter endpoint tree...' panel on the left lists various endpoints, with 'Genetic Toxicity' selected. The main table displays data for 9 chemical structures across various endpoints. A red callout '1' points to the 'Filter endpoint tree...' panel, and a red callout '2' points to the 'Genetic Toxicity' row in the table.

Structure	1	2	3	4	5	6	7	8	9
Structure	<chem>Oc1ccc(O)cc1</chem>	<chem>CCCCCCCC</chem>	<chem>CC(=O)C=C</chem>	<chem>Nc1ccc(O)cc1</chem>	<chem>CC(=O)O</chem>	<chem>CC(=O)O</chem>	<chem>CC(=O)O</chem>	<chem>CC(=O)O</chem>	<chem>CC(=O)O</chem>
Structure info									
Parameters									
Physical Chemical Properties									
Environmental Fate and Transport									
Ecotoxicological Information									
Human Health Hazards									
Acute Toxicity									
Bioaccumulation									
Carcinogenicity									
Developmental Toxicity / Teratogenicity									
Genetic Toxicity									
Immunotoxicity									
Irritation / Corrosion									
Neurotoxicity									
Photoinduced toxicity									
Repeated Dose Toxicity									
Sensitisation									
ToxCast									
Toxicity to Reproduction									
Toxicokinetics, Metabolism and Distribution									

1. **Remove** EC3 from the filter, click **Enter** 2. Position on the level of genetic toxicity as shown

Types endpoint correlations

Categorical vs. categorical
Gather experimental data – step 2

Example 2: Correlation of LLNA and AMES data

The screenshot displays the QSAR Toolbox interface during the 'Data' step. On the left, the 'Databases' panel shows 'Genotoxicity OASIS' selected with a green highlight (callout 1) and 'Genotoxicity ISSSTY' highlighted in red (callout 2). The 'Filter endpoint tree' on the right shows 'Genetic Toxicity' expanded, with 'Bacterial Reverse Mutation Assay' selected (callout 3). The main data matrix shows chemical structures in columns and assay results in rows, with 'M: Negative' and 'M: Positive' labels (callout 4).

Note that the correlation between endpoints is possible when data is gathered and available on data matrix. One should be aware of the data values that would be using during the data gap filling and gather the data for the corresponding endpoint during the "Endpoint" stage of the workflow, prior to entering the "Data gap filling" module

1. **Select** the databases including Ames data (green highlighted). Do not check ECHA Chem database.
2. Skin sensitization DB is already selected;
3. **Click** "Gather"
4. The data appeared on datamatrix;

Types endpoint correlations

Categorical vs. categorical
Define target endpoint – step 3

Example 2: Correlation of LLNA and AMES data

The screenshot shows the QSAR Toolbox interface with the following components:

- Top Bar:** QSAR TOOLBOX logo and navigation icons for Input, Profiling, Data, Category definition, Data Gap Filling, and Report.
- Left Panel:** Documents (Document 1, ToxCastDB, Skin Sensitization) and Data Gap Filling Settings (Only endpoint relevant, Only chemical relevant, At this position: Automated workflows, Standardized workflows).
- Filter endpoint tree:** A hierarchical tree on the left with 'LLNA' selected and highlighted in red. Other categories include Structure info, Parameters, Physical Chemical Properties, Environmental Fate and Transport, Ecotoxicological Information, Human Health Hazards, Genetic Toxicity, Immunotoxicity, Irritation / Corrosion, Neurotoxicity, Photoinduced toxicity, Repeated Dose Toxicity, and Sensitisation.
- Data Table:** A table with 7 columns and multiple rows. The row for 'LLNA' is highlighted in blue, and the cell containing 'M: Positive' is highlighted in blue. A blue callout bubble with the number '1' points to this cell.

Endpoint	1	2	3	4	5	6	7
Structure	<chem>O=C1C=CC(=O)N1</chem>	<chem>CCCCCCCC</chem>	<chem>CC(C)C(=O)CC</chem>	<chem>Nc1cccnc1</chem>	<chem>CC(C)C</chem>	<chem>CC(C)C</chem>	<chem>CC(C)C</chem>
Genetic Toxicity	M: Negative						M: Positive
Immunotoxicity							M: Positive
Sensitisation							
LLNA							M: Positive
EC3	M: Negative					M: Positive	M: Positive
A B C				M: Negative			
S W A N	M: Negative						
Undefined Assay	(1/1)						

The target endpoint is skin sensitization/in vivo/LLNA/EC3;
 1. **Click** on the cell associated with target endpoint;

Types endpoint correlations

Categorical vs. categorical

Enter Gap filling – step 4

Example 2: Correlation of LLNA and AMES data

1 Data Gap Filling

2 Read-across

3 Native scale/unit

4 OK

Converted data

Original data

Datapoints	#	Value	Original value	Assay
Human Health Hazards;Sensitisation	1	M: Positive (Skin sensitisation II (ECETOC))	1.06 % (Skin sensitization EC3(ratio))	LLNA

Enter Gap filling applying read across. Read across is applied because a categorical type data is analyzed.

1. Go to "Data Gap filling";
2. Select "Read-across";
3. Check "Skin sensitization I (OASIS)" scale;
5. Click "OK"

Types endpoint correlations

Categorical vs. categorical

Enter Gap filling – step 4

Example 2: Correlation of LLNA and AMES data

The screenshot shows the QSAR Toolbox interface during the 'Data Gap Filling' step. The 'Filter endpoint tree...' panel is expanded to show the 'Genetic Toxicity' endpoint, which is selected. The table below the tree shows the results of the gap filling process. An 'Information' dialog box is open, indicating that 13 observed values for 12 chemicals were excluded due to missing X descriptor value(s). A red '1' in a blue box points to the 'OK' button in the dialog. Below the screenshot, a text box explains the message and the required action.

The message informs the user for chemicals excluded from gap filling. 1. Click "OK";

Types endpoint correlations

Categorical vs. categorical

Perform correlation between GPMT and AMES data – step 5

Example 2: Correlation of LLNA and AMES data

The screenshot displays the QSAR Toolbox software interface during a workflow. The top menu bar includes 'Input', 'Profiling', 'Data', 'Category definition', 'Data Gap Filling', and 'Report'. The left sidebar shows 'Documents' and 'Data Gap Filling Settings' with options for 'Only endpoint relevant' and 'Only chemical relevant'. The central area features a 'Filter endpoint tree...' panel with a hierarchical view of endpoints like 'Neurotoxicity', 'Photoinduced toxicity', and 'Skin'. A 'Choose one' dialog box is open, showing radio button options for 'All', 'Mode', 'Lowest mode', 'Highest mode', 'Median', 'Lower median', 'Higher median', 'Minimal', and 'Maximal'. The 'Maximal' option is selected. Below the dialog is a scatter plot titled 'Read-across prediction for EC3, based on 7 values' with 'Observed: Positive; Predicted: Positive'. The plot shows data points for 'log Kow' (x-axis, -10 to 14) and 'EC3' (y-axis, Positive/Negative). A right sidebar contains a 'Calculation options' menu with 'Data usage' selected. Numbered callouts (1-4) indicate the steps: 1. Open 'Calculation' options; 2. Click on 'Data usage'; 3. Select 'Maximal'; 4. Click 'OK'.

1. **Open** "Calculation" options.
2. **Click** on "Data usage"
3. **Select** "Maximal"
4. **Click** "OK" (refer to slide 62 for details)

Types endpoint correlations

Categorical vs. categorical

Perform correlation between GPMT and AMES data – step 5

Example 2: Correlation of LLNA and AMES data

The screenshot displays the QSAR TOOLBOX interface during a 'Data Gap Filling' workflow. On the left, the 'Filter endpoint tree' shows a hierarchy of toxicity endpoints, with 'Sensitisation' and 'Genetic Toxicity' expanded. The central area features a data table with columns for 'target', 'log Kow', and 'M: Positive'. A 'Select endpoint descriptor' dialog box is open, showing a tree view of descriptors. A scatter plot at the bottom plots 'log Kow' (x-axis, -10 to 14) against 'M: Positive' (y-axis, Positive to Negative). Numbered callouts indicate the following steps: 1. Selecting 'Descriptors / data' in the 'Data Gap Filling Settings' panel; 2. Opening the 'Genetic Toxicity' node in the 'Filter endpoint tree'; 3. Selecting 'With S9' under 'In Vitro | Bacterial Reverse Mutation Assay (e.g. Ames Test) | Gene Mutation | Salmonella typhimurium'; 4. Clicking the 'OK' button in the 'Select endpoint descriptor' dialog box.

1. **Open** Select/descriptors data/ Select endpoint tree descriptor; 2. **Open** nodes under "Genetic Toxicity" node; 3. **Select** "With S9" under In Vitro|Bacterial Reverse Mutation Assay (e.g. Ames Test)|Gene Mutation| Salmonella typhimurium; ; 4. **Click** "OK" button;

Types endpoint correlations

Categorical vs. categorical

Perform correlation between GPMT and AMES data – step 5

Example 2: Correlation of LLNA and AMES data

The screenshot displays the QSAR Toolbox interface during a data gap filling workflow. A dialog box titled "Possible data inconsistency" is open, listing various endpoints and their associated data counts. A red "1" in a blue callout bubble points to the "OK" button in the dialog. The background shows a scatter plot of EC vs log Kow and a filter endpoint tree with "LLNA" selected.

Possible data inconsistency window content:

- No Strain Info (1 data; 1 chemicals)
- TA 100 (309 data; 176 chemicals)
- TA 102 (63 data; 59 chemicals)
- TA 104 (35 data; 34 chemicals)
- TA 1535 (239 data; 146 chemicals)
- TA 1537 (206 data; 130 chemicals)
- TA 1538 (133 data; 105 chemicals)
- TA 2638 (2 data; 2 chemicals)
- TA 7001 (6 data; 6 chemicals)
- TA 7002 (6 data; 6 chemicals)
- TA 7003 (5 data; 5 chemicals)
- TA 7004 (6 data; 6 chemicals)
- TA 7005 (6 data; 6 chemicals)
- TA 7006 (4 data; 4 chemicals)
- TA 97 (137 data; 102 chemicals)
- TA 97A (11 data; 11 chemicals)
- TA 98 (322 data; 191 chemicals)

Filter endpoint tree content:

- Immunotoxicity
- Irritation / Corrosion
- Neurotoxicity
- Photoinduced toxicity
- Repeated Dose Toxicity
- Sensitisation
 - Skin
 - In Vivo
 - GPMT (81/82)
 - HRIFT (80/125)
 - LLNA (601/713) M: Positive
 - EC3 (86/273)
 - Miscellaneous
- ToxCast
- Toxicity to Reproduction
- Toxicokinetics, Metabolism and Distribution

Possible data inconsistency window is appearing. **Click OK.**

Types endpoint correlations

Categorical vs. categorical

Perform correlation between GPMT and AMES data – step 5

Example 2: Correlation of LLNA and AMES data

The screenshot shows the QSAR Toolbox software interface during a data gap filling workflow. The main window displays a table of chemical structures and their predicted values for various endpoints. An information dialog box is open, stating "337 observed values for 294 chemicals were excluded due to missing X descriptor value(s)". A red circle with the number "1" highlights the "OK" button in the dialog box. The interface includes a menu bar, a toolbar, a left sidebar with document and settings panels, and a bottom panel with a scatter plot for EC3 prediction.

The appearing message inform for the common number gathered data across the number chemicals that will be excluded in Trend analysis due to missing X descriptor value(s). They are analogues with no AMES data. This will not affect the value of correlation coefficient; 1. **Click "OK";**

Types endpoint correlations

Categorical vs. categorical

Perform correlation between GPMT and AMES data – step 5

Example 2: Correlation of GPMT and AMES data

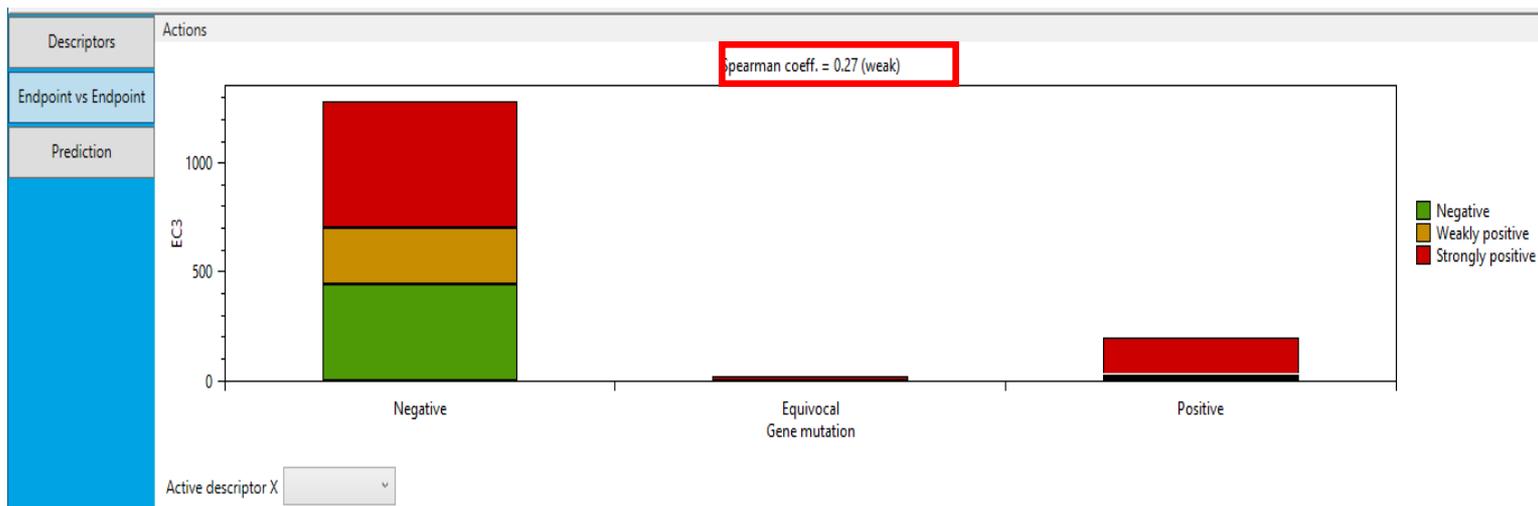
The screenshot displays the QSAR Toolbox software interface. The main window shows a data table with columns for chemical structures and rows for 'M: Negative' and 'M: Positive' values. An 'Edit calculator options' dialog box is open, showing a list of radio button options for calculating parameter values from a set of SMILES. The 'Maximal' option is selected. The dialog box also includes 'Additional options' and 'OK'/'Cancel' buttons. Below the table, a bar chart titled 'Spearman coeff. = 0.27 (weak)' shows the distribution of 'EC3' values for 'Negative', 'Equivalant Gene mutation', and 'Positive' categories. The chart has a legend with 'Negative' (green), 'Weakly positive' (yellow), and 'Strongly positive' (red). The interface also shows a 'Filter endpoint tree...' on the left and a 'Select / filter data' panel on the right.

1. Select "Descriptor/ data"; 2. Click "Edit descriptor options" ; 3. Select "Maximal" (worst case); 4. Click OK

Types endpoint correlations

Categorical vs. categorical

Interpretation of correlation results (GPMT vs. AMES)



Correlation analysis between two categorical type data: GPMT and AMES shows weak correlation between two endpoints (Spearman coefficient is 0.3).

Outlook

- Background
- Objectives
- The exercise
- **Workflow**
 - Correlation of data – background
 - **Types endpoint correlations**
 - Categorical vs. categorical
 - Categorized continuous vs. categorical

Types endpoint correlations

Categorized continuous vs. categorical

- The aim of this type correlation is to illustrate how categorized continuous and categorical type data correlates each other.
- Categorized continuous data is the continuous type data (e.g LC50 or AC50, EC3, %) converted into categories.
- In this example we will illustrate how DPRA ratio data (%) correlates with LLNA data:
 - DPRA (ratio data expressed in % and converted in categories)
 - LLNA (categorical type: Strongly positive, Weakly positive, Negative)
- Step by step workflow is presented on next few slides. Summary of the workflow steps are provided below:
 - *Load Skin sensitization database (step 1) – skipped, because this database has been already loaded on data matrix*
 - *Gather experimental data (step 2)*
 - *Define target endpoint (step 3)*
 - *Enter Gap filling (step 4)*
 - *Perform correlation between endpoints (step 5).*

Types endpoint correlations

Categorized continuous vs. categorical

Gather experimental data – step 2

Example: Correlation of DPRAs (%) and LLNAs (Strongly positive, Weakly positive, Negative) data

The screenshot shows the QSAR Toolbox interface. The top navigation bar has tabs for 'Input', 'Profiling', 'Data', 'Data Gap Filling', and 'Report'. The 'Data' tab is highlighted with a callout box labeled '3'. On the left, the 'Documents' panel shows a tree view with 'Skin Sensitization' selected, indicated by a callout box labeled '1'. Below it, the 'Databases' panel shows various property categories. The 'Filter endpoint tree...' panel on the left lists various endpoints, with 'In Chemico' highlighted by a red box and a callout box labeled '2'. The main area is a grid with 9 columns, each containing a chemical structure. The bottom row of the grid shows data for 'Acute Toxicity', 'Carcinogenicity', and 'Genetic Toxicity'.

1. **Click** again on the level of Skin sensitization; 2. Position the mouse on the level of In Chemico level of endpoint tree; 3. **Click** on Data tab

Types endpoint correlations

Categorized continuous vs. categorical

Gather experimental data – step 2

Example: Correlation of DPRA (%) and LLNA (Strongly positive, Weakly positive, Negative) data

The screenshot shows the QSAR Toolbox interface. The 'Data' menu is open, and the 'Gather' button is highlighted with a red circle and the number '3'. The 'Databases' section is expanded, and 'Chemical Reactivity COLIPA' is selected with a red box and the number '2'. The 'Documents' section shows 'Skin Sensitization' is selected. The 'Filter endpoint tree...' is visible on the left, and the 'Datamatrix' table is shown on the right. The table has 8 columns representing different chemical structures and rows representing various endpoints. Two rows are highlighted with red boxes and the number '5': 'In Chemo' and 'Sensitisation'.

Endpoint	1	2	3	4	5	6	7	8
In Chemo (314/772)			M: 12.5 %				M: 88 %	
Sensitisation (AW SW AOP (1201/2019))	M: Negative	M: Negative	M: Positive	M: Negative	M: Positive	M: Positive	M: Positive	M: Positive

1. Go to "Data"; 2. Select "Chemical reactivity COLIPA" database. Skin sensitization DB is already selected; 3. Click "Gather" button; 5. The data appeared on datamatrix;

Types endpoint correlations

Categorized continuous vs. categorical

Enter Gap filling – step 4

Example: Correlation of DPRA (%) and LLNA (Strongly positive, Weakly positive, Negative) data

Converted data

Original data

Datapoints	#	Value	Original value	Assay
Human Health Hazards;Sensitisation	1	M: Negative (Skin sensitisation II (ECETOC))	Negative (Skin sensitisation I (Oasis))	LLNA

Data points

Endpoint	Value	Assay
LLNA		
EC3	(614/727)	M: Negative
Miscellaneous	(419/801)	M: Negative
Undefined Assay	(1/1)	
ToxCast		
Toxicity to Reproduction		
Toxicokinetics, Metabolism and Distribution		

Native scale/unit

- Skin sensitisation I (Oasis)
- Skin sensitisation EC3(ratio)

Gap filling scale/unit

- Skin Sensitization (Danish ECETOC)
- Skin sensitisation I (Oasis)
- Skin sensitisation II (ECETOC)
- Skin sensitization EC3(ratio)
- Skin sensitization GHS (ordinal)

Information

13 observed values for 12 chemicals were excluded due to descriptor value(s)

Enter Gap filling and apply read across. Read across is applied because a categorical type data is analyzed.

1. Go to "Data Gap filling"; 2. Select "Read-across"; 3. Select "Skin sensitization I (OASIS)" scale ; 4-5. Click "OK";

Types endpoint correlations

Categorized continuous vs. categorical

Perform correlation between DPRA (%) and LLNA data – step 5

Example: Correlation of DPRA (%) and LLNA (Strongly positive, Weakly positive, Negative) data

The screenshot shows the QSAR Toolbox software interface. The top navigation bar includes 'Input', 'Profiling', 'Data', 'Category definition', 'Data Gap Filling', and 'Report'. The left sidebar contains 'Documents' and 'Data Gap Filling Settings'. The central workspace displays a 'Filter endpoint tree' on the left and a data table on the right. The data table has columns for 'Structure', 'Log Kow', and 'EC3'. A 'Choose one' dialog box is open over the data table, with a red '3' pointing to the 'Maximal' radio button. A red '4' points to the 'OK' button. The bottom right panel shows 'Calculation options' with a red '1' pointing to the 'Data usage' section and a red '2' pointing to the 'Use target data for prediction' option. A scatter plot at the bottom shows 'Read-across prediction for EC3, based on 5 values' with 'Predicted: Positive' and 'Log Kow' on the x-axis and 'EC3' on the y-axis. The y-axis categories are 'Strongly positive', 'Weakly positive', and 'Negative'. A blue box at the bottom contains the following instructions:

1. Open "Calculation options";
2. Click on "Data usage";
3. Select "Maximal"
4. Click "OK"

Types endpoint correlations

Categorized continuous vs. categorical

Perform correlation between DPRA and LLNA data – step 5

Example: Correlation of DPRA (%) and LLNA (Strongly positive, Weakly positive, Negative) data

The screenshot displays the QSAR Toolbox software interface. The main window shows a 'Select endpoint descriptor' dialog box with a tree view of descriptors. A red box highlights the 'DPRA (209/482)' descriptor under 'Physical Chemical Properties (306/760)'. A blue callout '3' points to this selection. Below the dialog, a 'Possible data inconsistency' dialog box is open, showing the selected endpoint and options for gap filling and data conversion. A blue callout '5' points to the 'Chemical reactivity DPRA 13% (ordinal)' option. A blue callout '6' points to the 'OK' button. A blue callout '7' points to the 'OK' button of the 'Information' dialog box. A blue callout '4' points to the 'OK' button of the 'Select endpoint descriptor' dialog box. A blue callout '1' points to the 'Descriptors / data' button in the right-hand sidebar. A blue callout '2' points to the 'Model/QSAR' button in the right-hand sidebar. A scatter plot at the bottom shows the correlation between DPRA and LLNA data, with a predicted positive correlation.

1. **Open Descriptors/data**
2. **Click** on "Select endpoint tree descriptor" button
3. **Click** on the endpoint tree on the level of "DPRA". In this case we mixed DPRA lysine and Cysteine data
4. **Click** on OK
5. **Select** Chemical reactivity DPRA 13% (ordinal) scale
6. **Click** OK
7. **Click** OK on the appeared message

Types endpoint correlations

Categorized continuous vs. categorical

Perform correlation between DPRA and LLNA data – step 5

Example: Correlation of DPRA (%) and LLNA (Strongly positive, Weakly positive, Negative) data

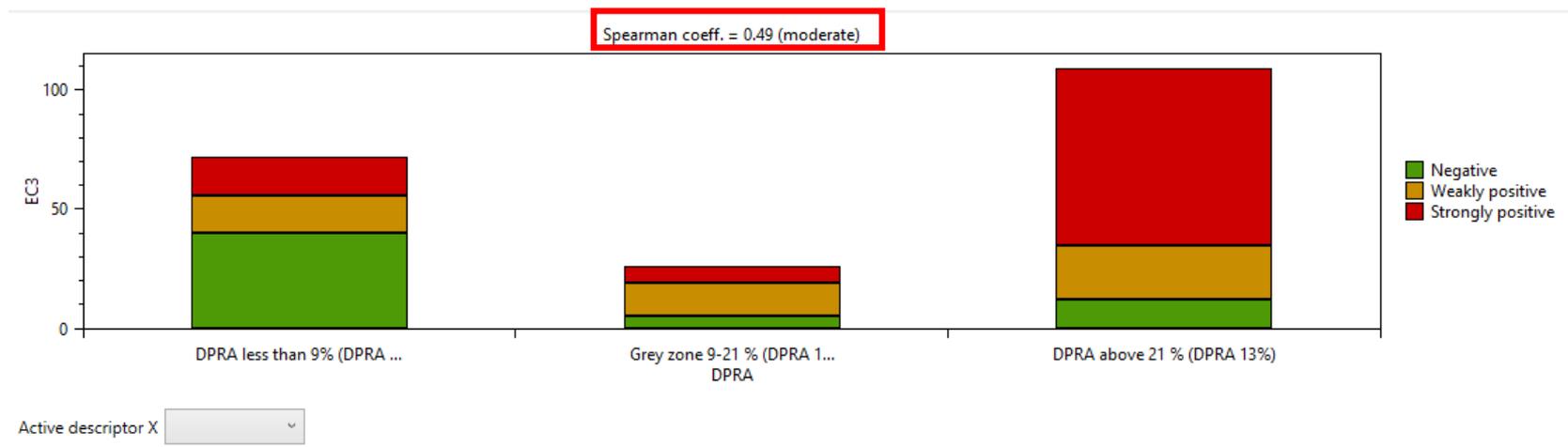
The screenshot shows the QSAR Toolbox interface during a correlation analysis. The top navigation bar includes 'Input', 'Profiling', 'Data', 'Category definition', 'Data Gap Filling', and 'Report'. The left sidebar lists various reports like 'Prediction Data Matrix', 'Category', and 'QMRF'. The main area displays a 'Filter endpoint tree' on the left and a data table on the right. The data table has columns for chemical structures and numerical values. A 'Prediction' panel at the bottom shows a stacked bar chart with three categories: 'DPRA less than 9% (DPRA ...)', 'Grey zone 9-21% (DPRA 1...)', and 'DPRA above 21% (DPRA 13%)'. The chart is labeled 'Spearman coeff. = 0.49 (moderate)'. A legend indicates 'Negative' (green), 'Weakly positive' (yellow), and 'Strongly positive' (red). Three callout boxes are present: 1. 'Open Edit descriptor options' (pointing to the 'Edit calculator options' dialog), 2. 'Select maximal values (worst case)' (pointing to the 'Maximal' radio button), and 3. 'Click OK' (pointing to the 'OK' button in the dialog).

1. **Open** Edit descriptor options
2. **Select** maximal values (worst case)
3. **Click** OK

Types endpoint correlations

Categorized continuous vs. categorical

Interpretation of correlation results (DPRA vs. LLNA)



- In this example we have correlate continues DPRA (%) data distributed into 3 bins (shown below) and categorical LLNA data (Strongly positive, Weakly positive, Negative)
 - Less than 9%
 - Grey zone 9 – 21%
 - Above 21%
- The high value of Spearman coefficient (0.49) shows moderate correlation between DPRA and LLNA data

Summary

- Different type correlations have been illustrated in this tutorial based on type of endpoint data:
 - Categorical vs. categorical:
 - Categorized continuous vs. categorical
- Correlation analysis has been evaluated by Spearman coefficient
- Moderate endpoint correlations have been obtained for 2 out of 3 illustrated examples.